International Journal of Pharmaceutical Research & Allied Sciences, 2019, 8(3):29-36



Research Article

ISSN : 2277-3657 CODEN(USA) : IJPRPM

Evaluation of Hepatoprotective Activity of Neem Extract in Rifampin Induced Acute Hepatic Failure in Rats

Maha A. Althaiban

Food and Nutrition Dept., Faculty of Home Economics, King Abdulaziz University, Jeddah, KSA.

ABSTRACT

The liver plays a role in many body functions such as immune defense and the metabolism of sugar and fat. Rifampin (RIF) is an antibacterial drug prescribed to treat tuberculosis (TB) along with multiple drugs. Other types of infections may also be treated with rifampin. Although the therapeutic effect of RIF has many adverse effects such as hepatoyoxicity. Neem (NM) has more than 140 compounds from various parts that have been isolated and can thus play a role in preventing hepatotoxicity. The research was carried out to examine the protective effect of neem leaves extract (NMLE) on RIF-induced liver damage. Forty male rats had been divided into four groups; Group I) non-treated negative control group, (Group II), which was given RIF (54 mg/kg/day) for thirty days, (groups III and IV intoxicated rats received orally the NMLE in doses of two hundred and fifty and five hundred mg/kg/day respectively, for 30 days. At day 30, blood was collected for biochemical analysis, as well as the liver was also examined histopathologically. The results revealed that the NMLE at the two dosage levels significantly decreased serum levels of liver enzymes and MDA accompanied by significantly increased in activities of GSH, SOD, and showed ant-inflammatory effects as evidence by significantly decreased in TNF-a and IL-1a levels compared to RIF group II. There was also an improvement in histopathological alterations observed in liver tissues of hepatotoxic rats. Therefore, the administrations of NMLE has hepatoprotective effects in hepatotoxic rats via antioxidant and ant-inflammatory pathway.

Key words: Liver toxicity, Neem, Rifampin, Rats, Cytokines, Histopathological.

INTRODUCTION

Liver is an important organ in metabolism of drugs. Drug that induce liver failure is a major medical problem [1]. Tuberculosis, an immunodeficiency virus, is one of a major problems worldwide. It is from the deadliest causes among curable infectious diseases, caused in human by various strains of mycobacteria; Mycobacterirum tuberculosis, MTB. It is spread through the air from transmitting the saliva of people who have an active MTB infection that attacks lungs [2, 3]. Rifampicin (RIF), an anti-tuberculosis drugs, is an antibiotic produced by stretomyces mediterrenei that induced hepatocellular dysfunction with early stage of administration [4, 5]. It affects bilirubin excretion and induced hyperbilirubinemia [6], and causes cholestasis, hepatic lesions, centrilobular necrosis and liver injury [6, 7].

Production of ROS is a crucial factor inducing tissues oxidative damage [8]. Herbal medicine has gained attention for maintaining health [9]. It plays a major role in the management of liver' disorders via regeneration of liver cells and prevention the oxidative damage induced by ROS [10].

The features of plants have created world-wide interest about medicinal plants which lead to novel sources of drugs for wide modern applications [11]. They are also widely utilized to control various insect pests [12]. *Azadirachta indica*, known as Neem, is therapeutic plant. It possesses diverse pharmacological properties. Neem leaves extract has anti-inflammatory, antipyretic and antimicrobial activities [13-15]. It showed hyperglycemic and hypolipidemic activities in diabetic rats [16], and gastroprotective properties [17]. Neem leaf comprises several valuable constituents as polyphenolics, flavonoids, glycosides, querctin and terpenoids, which possess gastroprotective [18, 19]. Administration of NMLE protected liver from cisplatin hepato- and nephro-toxicity

[20]. This work aims to assess the protective role of NMLE on RIF-induced hepatotoxicity.

MATERIALS AND METHODS

Plant material

Neem (A. indica) (NM) leaves, were bought from local market, Jeddah, KSA.

Drugs and chemicals

Rifampin (RIF) was purchased from a local pharmacy (Jeddah). All chemicals and kits with high analytical grade were bought from Sigma-Aldrich Co., USA.

Rats

Sprague Dawley adult male rats $(180\pm 10 \text{ g})$ were provided from King Fahd Medical Research Center (KFMRS), KAU. They were kept in standard laboratory conditions, fed on a standard AIN-93 diet [21]. They were kept in accordance to the standard guidelines in KFMRS, KAU for the use and care of laboratory animals.

Plant material and extraction

Five hundred grams of the NM fresh leaves were cleaned with distilled water, then soaked in 500 ml ethanol (80%), for 48 h at room temperature. The extract was filtrated, evaporated by a rotary evaporator, then concentrated on water bath (36° C). The extract, greenish paste, stored at ($<4^{\circ}$ C) until use [22].

Protocol design

Wister male rats (n=40, 180 \pm 10 g) were used in the experiment. The rats fed standard formula diet in a steady environment (22 -25 °C temp., 45-55% humidity). Rats were put under the observation for a week before the beginning of the protocol experiment. After the acclimatization period, the rats were randomly distributed to 5 groups (ten each).

Group I: Control negative(Cont); rats administered orally1 ml/kg (1% CMC) for 30 d.

Group II: Control positive (RIF); rats treated orally with RIF (54 mg/kg/day) for 30 d. [23].

Group III: Rats received orally NME (250 mg/kg/day) [24], and receive RIF 1 h after administration of NMLE for 30 d.

Group IV: Rats received orally NME (500 mg/kg/day) and receive RIF 1 h after administration of NMLE for 30 d

After 30 d of experimental time, all rats for each group sacrificed under light anesthesia with ether. Blood samples were collected by heparinized capillary tubes, kept for couple of h and centrifuged for 15 min at 3000 rpm. Then serum samples stored at -20° C for subsequent followed analyzes. The liver was removed for histopathological examination.

Determination of serum biomarkers

Serum liver enzymes activities (aspartate aminotransferase (AST), alanine aminotransferase (ALT)) determined as described [25], and alkaline phosphatase (ALP) [26]. Oxidative stress biomarkers (reduced glutathione (GSH), superoxide dismutase (SOD), and malondialdehyde (MDA)) tested chemically [27-29], respectively]. Anti-inflammatory cytokines (interleukin– 1α (IL- 1α) and tumor necrosis factor- α (TNF- α)) were tested [30, 31, respectively].

Histopathological examination

The liver tissue samples from all groups were prepared and stained with hematoxylin-eosin by routine procedures [32].

Statistic

The analysis of the results was done by SPSS ver. 24, one-way ANOVA. Values were showed as mean \pm SDM. The results considered significance at $P \le 0.05$ [33].

RESULTS

Administration of RIF orally to rats caused significant (P < 0.05) decreases in BWG %, FI and FER (24.18 ± 0.43,18.88 ± 0.089 and 0.013 ± 0.009, respectively) when compared to Cont group (64.75 ± 0.61, 31.22 ± 0.32 and 0.021 ± 0.004, respectively). Oral administration of NMLE to rats inflicted with hepatoxicity induced significant (P < 0.05) increases in all biological parameters as compared with RIF group (Table 1).

Table 1. Hepatoprotective effect of Neem leaves extract (NMLE) on biological evaluation in hepatotoxic-rats.

Groups	BWG %	FI (g/rat/day)	FER
Cont	$64.75\pm0.61a$	31.22 ± 0.32 a	0.021 ± 0.004 a

RIF	$24.18\pm0.43~b$	$18.88\pm0.089~b$	$0.013 \pm 0.009 \text{ b}$
NMLE (250 mg/kg)+RIF	59.38 ± 1.15 a	28.56 ± 0.65 a	0.021 ±0.001a
NMLE (500 mg/kg)+RIF	61.53 ± 1.26 a	29.76 ± 0.97 a	0.021 ± 0.005 a

- Values are presented as mean \pm SDE.

- In the same column different superscript letters considered significantly different (P < 0.05).

From data recorded in Table (2) it might be recognized that intoxicated rats by RIF (54 mg/kg) caused significant (P<0.05) increases the serum liver enzyme activities (AST, ALT and ALP: 141.19±2.13, 130.36±0.26 and 143.22±2.43, respectively) as compared to Cont group (53.56±1.65, 48.21± 0.53 and 58.92±2.15, respectively). Administration of NMLE (250 and 500 mg/kg) to intoxicated with a RIF exhibited significant decreases (P<0.05) in all the elevated liver markers as compared with RIF group.

The NMLE effect on MDA, GSH and SOD in hepatotoxic rats is shown in Table (3). In a group of hepatotoxicity, MDA increased significantly (25.37 ± 1.32) compared to Cont rats (13.34 ± 1.54), while there were significant decreases in GSH and SOD (3.87 ± 0.09 and 23.46 ± 1.45 , respectively) compared to Cont rats (13.34 ± 1.54 , $6.27\pm.03$ and 57.34 ± 1.95 , respectively). The pretreatment with NMLE at the two dosage levels resulted in significant increases in MDA level and significant decreases in GSH and SOD activities compared to RIF hepatotoxic rats. Pretreatment with NMLE at a dose of 500 mg/kg/ day showed strong efficacy in MDA, GSH and SOD compared to pre-treated group a dose of 250 mg/kg.

Anti-inflammatory cytokines IL-1 β and TNF- α serum levels are significantly (P < 0.05) increased in rats intoxicated by RIF (54 mg/kg) compared with their values in the Cont group. Administration of NMLE extract in doses of 250 and 500 mg/kg/ day results in a significant decrease in serum IL-1 β and TNF- α compared to hepatotoxic group. Administration of NMLE extract in a dose of 500 mg/kg results in non-significant serum IL-1 β and TNF- α compared to Cont group. The high dose (500mg/kg) of NMLE extract is the most effective compared to the NMLE low dose group as presented in Table (4).

Table 2.	Hepatoprotective effect of Neem	leaves extract	(NMLE) o	on serum	liver enzy	mes AST, AL	T and ALP
		in hepatoto	xic rats.				

Groups	AST (U/L)	ALT(U/L)	ALP (U/L)
Cont	53.56±1.65 c	48.21± 0.53 c	58.92±2.15 c
RIF	141.19±2.13 a	130.36±0.26 a	143.22±2.43a
NMLE (250 mg/kg)+RIF	67.53±2.67 b	75.37±2.85 b	81.32±1.45a
NMLE (500 mg/kg)+RIF	56 .20±3.43 c	54.64±0.62 c	62.59±2.21 b

- Values are presented as mean \pm SDE.

- In the same column different superscript letters considered significantly different (P<0.05).

 Table 3. Hepatoprotective effect of Neem leaves extract (NMLE) on the levels of MDA, GSH and SOD in hepatotoxic rats

	1		
Groups	MDA	GSH	SOD
Groups	(nmol/g protein)	(µg/mg protein)	(U/mg)
Cont	13.34±1. 54 c	6.27±.0.03a	57.34 ± 1.95 a
RIF	25.37±1. 32 a	3.87±0.09 c	23. 46 ± 1.45 c
NMLE (250 mg/kg)+RIF	19.36±1.32 b	5.07±0.05 b	64.23±1.74 b
NMLE (500 mg/kg)+RIF	14 .30±1.12 c	6.14±0.08 a	59.03±1.12 a

- Values are presented as mean \pm SDE.

- In the same column different superscript letters considered significantly different (P < 0.05).

Table 4. Hepatoprotective effect of Neem leaves extract (NMLE) on serum IL-1 and TNF- α level in hepatotoxic rats

Groups	IL-1 (pg/ml)	TNF-α (pg/ml)		
Cont	23.82 ± 0.24 c	$12.03 \pm 1.26 \text{ c}$		
RIF	58.41± 0.21 a	27.53 ± 0.74 a		
NMLE (250 mg/kg)+RIF	33.26±1.30 b	19.49± 0.06 b		
NMLE (500 mg/kg)+RIF	24 .76±1.63 c	14.84±0.03 c		

- Values are presented as mean ± SDE.

- In the same column different superscript letters considered significantly different (P < 0.05).

Liver tissue of Cont rats showed normal histological structure of hepatic lobule with normal hepatic sinusoids

and hepatocytes (Fig.1.A). Administration of RIF to rats induced inflammatory cell infiltration, hepatic necrosis and hepatocytes apoptosis (Fig.1.B). Liver tissue of rats given NMLE (250mg/kg) + RIF showed slight congestion of hepatic sinusoids (Fig.1.C). In group received NMLE (500 mg/kg) + RIF revealed apparently normal architecture of hepatic tissue (Fig.1.D).



Figure 1: Hepatoprotective effect of Neem leaves extract (NMLE) on liver sections in hepatotoxic rats (H&E stain x 400). Liver section of Cont rats (-ve) showed normal histological structure of hepatic lobule [A]. In RIF intoxicated rats' liver section showed inflammatory cell infiltration, focal hepatic necrosis and apoptosis of hepatocytes [B]. Liver section of pretreated intoxicated rats with NMLE (250mg/kg) showed slight congestion of hepatic sinusoids [C]. Liver section of pretreated intoxicated rats with NMLE [500 mg/kg] showed normal hepatic lobule.

DISCUSSION

Rifampin is one of the major anti-TB drugs prescribed in the treatment of mycobacterial infections. However, it has adverse effect, where it is a leading cause of acute liver failure, which is mainly related to oxidative stress [34, 35]. Neem is versatile medicinal herbs having many biological activities [36]. Its phytochemical constituents have anti-inflammatory, antimicrobial, antioxidant and diverse pharmacological activities [15, 16]. This study aimed to assess the potential protective role of NMLE against RIF-induced acute liver injury in rats. In this work, administration of RIF caused significant decrease in biological evaluation compared to the negative control group. This effect of RIF explained via its negative impact engendered through oxidative stress [35, 37]. While the non-significant changes in biological evaluation were observed in the groups administrated NMLE (250 and 500 mg/kg) + RIF compared with Cont group, which reflects the inhibitory effect of NMLE against RIF. The same findings were reported by Nwobodo et al. [38] who found that there was a significant increase of body weight in the group pretreated with *A. indica* and paracetamol compared with once treated only with paracetamol, thus could explained via the inhibitory effect of *A. indica* on the oxidative stress induced by paracetamol.

Liver enzymes transaminases and alkaline phosphatase activities show the liver state [39]. RIF group revealed significant increases in the liver enzyme activities comparing with negative control. This is considered a sign of hepatic intoxication and damage [40, 40]. This could be explained via the hepatocellular damaged by RIF induced releasing of enzymes located in the cytosol into the blood stream [42]. In addition, RIF produced cytochrome P450 enzyme lead to elevated production of toxic metabolites from acetyl hydrazine [43]. Hepatoprotective activity of NMLE (250 and 500 mg/kg)+ RIF have been proved via the enhanced liver enzyme activities compared to the RIF group. This results are in agreement with Kausik et al. [36] and Adeshina et al. [44]. Thus, it could be explained by inhibition peroxidation effect by NMLE, which maintains liver's cell membrane integrity [17, 45].

RIF significantly induced oxidative stress and inflammation via significant increase the levels of MDA, as well as IL-1 β and TNF- α , with significant deplete in the antioxidant enzymes GSH and SOD activities compared to Cont rats. The observed results agree with Yuhas et al. [46] and Mlambo and Sigola [47] who reported that RIF

increases the cytokines production IL-1 β and TNF- α by increasing iNOS mRNA transcription and iNOS protein expression. Anti-TB drugs induced hepatotoxicity via oxidative stress through stimulation of lipid peroxidation as a source of cells membrane destruction and damage, and diminution of cellular defense antioxidant mechanisms which metabolizes toxic compounds to non-toxic compounds [42, 48-50].

Pretreatment with NMLE has anti-inflammatory effect and restored the antioxidant status in a dose dependent manner. This could be due to the active components of NMLE such as flavonoids, nimbin, 6-desacetylnimbinene, quercetin, nimbandiol, 17-hydroxy azadiradione, ascorbic acid, nimbolide, n-hexacosanol, 7-sdesacetyl-7- benzoylazadiradione,7-sdesacetyl-7-benzoylgedunin, and nimbiol [51, 52], that have anti-inflammatory, immunomodulating and antioxidant properties, thereby preventing the oxidative stress and inflammation imposed by RIF, which are the most important mechanisms in hepatotoxicity of this drug [14, 53, 54].

The RIF induced focal hepatic necrosis, inflammatory cell infiltration and apoptosis of hepatocytes. RIF induced liver necrosis [55], thus established in elevated levels of AST and ALT, which used to assess hepatocellular damage and leading to liver cell necrosis [56]. Pretreatment with NMLE almost normalize liver histological architecture. This finding is found in several studies [57-59], thus could be attributed to the phytochemical constituents in NMLE which have hepatoprotective activity [14].

It could be concluded that NMLE attenuates the hepatic damage induced by RIF, it prevents all biological and biochemical parameters, as well as histopathological alterations via antioxidant and anti-inflammatory pathways.

REFERENCES

- 1. Holt, M.; Ju, C. Drug-induced liver injury. Handb. Exp. Pharmacol. 2010, 196, 3–27.
- 2. Kumar, V., Abbas, A.K., Fausto, N. and Mitchell, R.N. (2007). Robbins Basic Pathology, 8th ed., Saunders Elsevier. pp. 516-22.
- 3. Konstantinos, A. (2010). Testing for tuberculosis, Australian Prescriber 33 (1): 12-18.
- 4. Geo,F.B., Karen, C.C.,Butel, J.S., Morse, S.A. and Timothy, A.M (2010). Melnick and Alderberg"s medical microbiology: McGrawhill.
- 5. Haddad, L. and Winchester, J. (1983). Clinical Management of Poisoning and Drug Overdose. WB Saunders Co.
- 6. Danan, G. and Teschke, R. (2016). RUCAM in drug and herb induced liver injury: The update. Int. J. Mol. Sci., 17:14.
- 7. Singh, J., Arora, A., Garg, P., Thakur, V., Pande, J. and Tandon, R. (1995). Antituberculosis treatment-induced hepatotoxicity: Role of predictive factors. Postgrad. Méd. J. 1995, 71, 359-62.
- 8. Dash, D. K., Yeligar, V. C. and Nayak, S. S. (2007). Evaluation of hepatoprotective and antioxidant activity of Ichnocarpus frutescens (Linn.) R.Br. on paracetamol-induced hepatotoxicity in rats. Tropical Journal of Pharmaceutical Research, 6 (3):755-65.
- 9. Bent, S. (2008). Herbal medicine in the United States: review of efficacy, safety, and regulation: grand rounds at University of California, San Francisco Medical Center. Journal of General Internal Medicine, 23(6):854-9.
- 10. Kucharska, J., Uli ' cn a, O., Gvozdj ' akov ' a A. (2004). Regeneration ' of coenzyme Q9 redox state and inhibition of oxidative stress by Rooibos tea (Aspalathus linearis) administration in carbon tetrachloride liver damage. Physiological Research, 53(5): 515-21.
- 11. Ramakrishna, N., Saidulu, Ch. Medicinal Plants Used By Ethnic People of Adilabad District, Andhra Pradesh, India. Int. J. of Pharm. Res. & All. Sci., 2014, 3(2), 51-59.
- Ojewumi, M.E., Adedokun, S.O., Omodara, O.J., Oyeniyi, E.A., Taiwo, O.S., Ojewumi, E.O. Phytochemical and Antimicrobial Activities of the Leaf Oil Extract of Mentha Spicata and its Efficacy in Repelling Mosquito. International Journal of Pharmaceutical Research & Allied Sciences, 2017, 6(4), 17-2.
- 13. Stirnimann, G., Kessebohm, K. and Lauterburg, B. (2010). Liver injury caused by drugs: An update. Swiss Méd. Wkly., 140, 18.
- 14. Kale, B.P., Kothekar, M.A., Tayade, H.P., Jaju, J.B. and Mateenuddin, M. (2003) Effect of aqueous extract of Azadirachta indica leaves on hepatotoxicity induced by antitubercular drugs in rats. Indian Journal of Pharmacology; 35: 177-80.

- 15. El-Hawary,S.S., El-Tantawy,M.E., Rabeh,M.A. and Badr,W.K. (2013). Chemical composition and biological activities of essential oils of Azadirachta indica. A. Juss, International Journal of Applied Research in Natural Products, 6: 33-42.
- 16. Prashanth, G.K. and Krishnaiah, G.M. (2014). Chemical composition of the leaves of Azadirachta Indica Linn (Neem). Int. J. Adv. Eng. Tech. Manag. and Appl. Sci. 1(5): 21-31.
- 17. Chattopadhyay, R.R. and Bandyopadhyay, M. (2005). Effect of Azadirachta indica leaf extract on serum lipid profile changes in normal and streptozotocin induced diabetic rats. African Journal of Biomedical Research, 8; 101-4.
- Bandyopadhyay, U., Biswas, K., Chatterjee, R., Bandyopadhyay, D., Chattopadhyay, I., Ganguly, C.K., Chakraborty, T., Bhattacharya, K. and Banerjee, R.K. (2002). Gastroprotective effect of Neem (Azadirachta indica) bark extract: possible involvement of H(+)-K(+)-ATPase inhibition and scavenging of hydroxyl radical. Life Sci., 71: 2845-65.
- Boeke, S.J., Boersma, M.G., Alink, G.M., van Loon, J.J., van Huis, A., Dicke, M. and Rietjens, I.M. (2004). Safety evaluation of neem (Azadirachta indica) derived pesticides. J. Ethnopharmacol., 94: 25-41.
- 20. Sarkar, K., Bose, A., Laskar, S., Choudhuri, S.K., Dey, S., Roychowdhury, P.K. and Baral, R. (2007). Antibody response against neem leaf preparation recognizes carcinoembryonic antigen. Int. Immunopharmacol., 7: 306-12.
- 21. Reeves, P.G., Nielsen, F.H. and Fahey Jr., G.C. 1993. AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76A rodent diet. Journal of Nutrition., 123 (11):1939–51.
- 22. Efiong, E.E., Igile, G. O., Mgbeje, B.I.A., Out, E.A. and Ebong, P.E. (2013) Hepatoprotective and antidiabetic effect of combined extracts of M. oleifera oleifera and Vernonia amygdalina in streptozotocininduced diabetic albino Wistar rats, Journal of Diabetes and Endocrinology, 4(4):. 45-50.
- 23. Swamy ,V., Rucha, V., Kulkarni, B., Koti, P. C. and Gadad, A. H. M.(2012) Hepatoprotective effect of cissus quadrangularis Stem extract against rifampicin-induced Hepatotoxicity in rats, Indian Journal of Pharmaceutical Sciences, 74 (2): 183-7.
- Shravan, K.D., Ramakrishna, R., Santhosh, K.M., and Kannappan, N. (2011) In vivo Antidiabetic evaluation of Neem leaf extract in alloxan induced rats. Journal of Applied Pharmaceutical Science., 1(4): 100-5.
- 25. Reitman, S. and S. Frankel, 1957. A colorimetric method for the determination of serum oxaloacetic and glutamic pyruvic transaminases. Am. J. Clin. Pathol., 28:56.
- 26. Belfied, A. and Goldberg, D.M. (1971). Enzyme, 12,561. C/F: Bio-Merieux, L'Etoile, France T. Yoshioka, K.Kawada, T. Shimada, and M.Mori, "Lipid peroxidation 44-52.
- 27. Giannopolitis, C. N. and Ries, S. K. (1977). Superoxide dismutase. I. occurrence in higher plants. Plant Physiology, 59(2): 309-14.
- 28. Spitz, D.R. and Oberley, L.W. (1989). An assay for superoxide dismutase activity in mammalian tissue homogenates. Anal. Biochem., 179: 8-18.
- 29. Beutler, E., Duron, O. and Kelly, B.M. (1963). Improved method for the determination of blood glutathione. J. of Laborat. and Clin. Med.,61:882-8.
- Piguet, P.F., Grad, G.E., Allet, B. and Vassalli, P. (1987). Tumor necrosis factor/cachectin is an effector of skin and gut lesions of the acute phase of graft-versushost disease. J. Exp. Med., 166:1280-89
- Dinarello, C. A. (2009). Immunological and inflammatory functions of the interleukin-1 family. Annu. Rev. Immunol., 27, 519-50
- 32. Bancroft, D., Stevens, A. and Turmer, R. (1998). Theory and practice of histological technique, 4th ed., Churchill Living Stone, Edinburgh, London, Melbourne. 47-67.
- 33. Snedecor, G.W. and Cochron, W.G. (1989). Statistical methods. 8th edit. Lowa State Univ. Press, Ames, Lowa, USA.
- Devarbhavi, H., Singh, R., Patil, M., Sheth, K., Adarsh, C.K. and Balaraju, G. (2013). Outcome and determinants of mortality in 269 patients with combination anti-tuberculosis drug-induced liver injury. J. Gastroenterol. Hepatol, 28:161-7.

- 35. Huang, J.H., Zhang, C., Zhang, D.G., Li, L., Chen, X. and Xu, D.X. (2016). Rifampicin-induced hepatic lipid accumulation: Association with up-regulation of peroxisome proliferator-activated receptor gamma in mouse liver. PLoS ONE, 11, e0165787.
- 36. Kausik B, Ishita C, Ranajit KB, Uday B. 2002. Biological Activities and Medicinal Properties of Neem (Azadirachta indica). Current Sci., 82(11): 1336-45.
- Chowdhury, A., Santra, A., Bhattacharjee, K., Ghatak, S., Saha, D.R. and Dhali, G.K. (2006). Mitochondrial oxidative stress and permeability transition in isoniazid and rifampicin induced liver injury in mice. J. Hepatol., 45, 117–26.
- Nwobodo, E. I., Nwosu, D. C., Ogbodo,S.O., Ugwuene, F. O., Ihim,A.C., Onuabuchiani, N., Nnodim, J. K. and Ani, O. (2018). Effects of Azadirachta indica leaf aqueous extract on the antioxidant enzymes in paracetamol-induced hepatotoxicity in Wistar rats. Int. J. Biol. Chem. Sci. 12(1): 1-10.
- 39. Suja, S.R., Latha, P.G., Pushpangadan, P. and Rajasekharan, S. (2004). Evaluation of hepatoprotective effects of Helminthostachys zeylanica hook against carbon tetrachloride-induced liver damage in Wistar rats. Journal of Ethnopharmacol., 92: 61-6.
- 40. Adeyemi, O.S. and Akanji, M.A. (2011). Biochemical changes in the kidney and liver of rats following administration of ethanolic extract of Psidium guajava leaves. Human and Experimental Toxicol., 30(9): 1266-74.
- Kpemissi, M., Metowogo, K., Lawson-Evi, P., Eklu-Kadégbékou, K., Aklikokou, A.K. and Gbéassor, M. (2015). Hepatoprotective and antioxidant effects of Acanthospermum hispidum (DC) leaves on carbon tetrachloride-induced acute liver damage in rat. Int. J. Biol. Chem. Sci., 9(5): 2263 -71.
- 42. Anbarasu, C., Rajkapoor, B. and Kalpana, J. (2011). Protective effect of Pisonia aculeata on rifampicin and Isoniazid induced hepatotoxicity in rats. International Journal of Phytomedicine 3: 75-83.
- 43. Ramaiah, S.K., Apte, U. and Mehendale, H.I.M. (2001). Cytochrome P4502E1 induction increases thioacetamide liver injury in diet-restricted rats. Drug Metab. Dispos., 29:1088-95.
- 44. Adeshina, A.J., Fakunle, P.B. and Oloyede, A.O. (2011). Some protective effects of aqueous leaf extract of Azadirachta indica on paracetamol-induced hepatotoxicity in adult Wistar rats. Am J. of Tropical Med. Public Heal., 1(3): 97-106.
- 45. Nwobodo, E.I. (2017). Evaluation of antilipid peroxidation and hypolipidemic potentials of Azadirachta indica leaf aqueous extract in paracetamol induced hepatotoxicity in Wistar rats. Int. J. Inform. Res. Rev.,4(2): 3615-9.
- 46. Yuhas, Y., Berent, E. and Ashkenazi, S. (2011). Effect of Rifampin on production of inflammatory mediators in HepG2 liver epithelial cells. Antimicrobial Agents and Chemotherapy, 55(12): 5541–6.
- 47. Mlambo, G. and Sigola. L. B. (2003). Rifampicin and dexamethasone have similar effects on macrophage phagocytosis of zymosan, but differ in their effect on nitrite and TNF- production. Int. Immunopharmacol. 3:513-22.
- 48. Mitra, S.K., Venkataranganna, M.V., Sundaram, R. and Gopumadhavan, S. (1998). Protective effect of HD-03, a herbal formulation, against various hepatotoxic agents in rats. J Ethnopharmacol.,63:180-6.
- 49. Sodhi, C.P., Rana, S.F., Attri, S., Mehta, S., Yaiphei, K. and Mehta, S.K. (1998). Oxidative-hepatic injury of isoniazid-rifampicin in young rats subjected to protein and energy malnutrition. Drug Chem Toxicol., 21:305-17.
- 50. Georgieva, N., Gadjeva, V. and Tolekova, A. (2004). New isonicotinoylhydrazones with ssa protect against oxidativehepatic injury of isoniazid. TJS;2:37–43.
- 51. Kokate, C., Purohit, A.P. and Gokhale, S.B. (2010). Pharmacognosy. Nirali Prakashan India, 10: 28-29.
- 52. Hossain, M.A., Al-Toubi, W.A.S., Weli, A.M., Al-Riyami, Q.A. and Al-Sabahi JN. (2013). Identification and characterization of chemical compounds in different crude extracts from leaves of Omani neem. J. Taibah Univ. Sci., 7(4): 181-8.
- Sodhi, C.P., Rana, S.V., Mehta, S.K., Vaiphei, K., Attri, S. and Thakur, S. (1996). Study of oxidative stress in isoniazid induced hepatic injury in young rats with and without protein energy malnutrition. J Biochem Toxicol., 11:139-46.
- 54. Sodhi, C.P., Rana, S., Mehta, S., Vaiphei, K., Goel, R.C. and Mehta, S.K. (1997). Study of oxidative stress in rifampicin induced hepatic injury in growing rats with and without protein energy malnutrition. Hum Exp Toxicol., 16:315-21.
- 55. Hussain, Z., Kar, P. and Husain, S.A. (2003). Antituberculosis drug-induced hepatitis: Risk factors, prevention and management. Indian J Experimental Biol., 41:226.

- 56. Amacher, D.E. (1998). Serum transaminase elevations as indicators of hepatic injury following the administration of drugs Regul Toxicol Pharmacol., 27:119-130.
- 57. Bhanwra, S, Singh, J. and Khosla, P. (2000). Effect of Azadirachta indica (Neem) leaf aqueous extract on paracetamol-induced liver damage in rats. Indian J. Physiol. Pharmacol., 44: 64-8.
- 58. Yanpallewar, S.U., Sen, S., Tapas, S., Kumar, M., Raju, S.S. and Acharya, S.B. (2003). Effect of Azadirachta indica on paracetamol-induced hepatic damage in albino rats. Phytomedicine, 10: 391-6.
- Boeke, S.J., Boersma, M.G., Alink, G.M., van Loon, J.J., van Huis, A., Dicke, M. and Rietjens, I.M. (2004). Safety evaluation of neem (Azadirachta indica) derived pesticides. J. Ethnopharmacol., 94: 25-41.