



Research Article

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Parasitological, bacteriological and in-vitro studies on antibacterial activity of ethanolic extract of Calligonumcomosum in goats affected with fascioliasis in Taif, KSA.

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ABSTRACT

This study was carried out to detect the relationship between bacterial infections and liver fascioliasis. Grossly, post mortem examination of the livers infested with Fasciolagigantica in the present study, were hard, firm and tough in consistency with multiple, irregular pale brownish areas on the surface. Cut section of the liver revealed presence of large whitish areas of fibrosis. The affected ducts were enlarged, thickened, hard in consistency and in the form of cord like structures and protruded above the surface of the liver. In most cases, mature Fasciola worms were detected within the lumen of the affected bile ducts and bile stones. Several types of bacteria were isolated from liver surface, bile duct and Fasciola worms from goats suffering from chronic fascioliasis. Bacterial isolated from liver surface were Stenotrophomonasmalyophilia, Corynebacteriumpropinquum and Escherichia coli. The bacteriological bile culture, revealed isolation of AerococcusUrinae and Escherichia coli. While bacterial species isolated from Fasciolagigantica, were Leifsonia aquatic, Corynebacteriumrenale group, Corynebacterium species, Escherichia coli and Lactococcuslactis spp. cremoris. All the isolated microorganisms are pathogenic and can transmitted from animals to human causing very dangerous zoonotic diseases. The present investigation evaluated the in-vitro antibacterial activity of the crude ethanolic extracts of Calligonumcomosum 10% on some gram positive and negative bacteria. Further in vivo studies are required to validate our findings and improve our knowledge on the potential of the Calligonumcomosum extract as antimicrobial in relation to its chemical composition.

Keywords:*Fasciola ,Calligonumcomosum , goats, antibacterial.*

INTRODUCTION

Fasciola species plays an important role in the microbial invasion of the infected animals either by transportation or depressing the vital resistance of the host. Besides, infected liver constitutes a good media for bacterial multiplication, transportation of micro-organisms with the parasites occurs during the different stages of its life cycle either outside or inside the animal body. Anaerobic necrotic lesions of the liver produced by immature flukes occasionally provides a suitable environment for the germination of spores of Clostridium novyi type B bacteria in the liver. The bacteria will release toxins into the blood stream resulting in what is known as black disease in sheep and goats and sometimes cattle (1), (2) and (3).

Control of fascioliasis may be achieved by control of snail and by use of anthelmintic treatment of infected animals, (4). Snail control is difficult and expensive, and involves improved drainage, fencing and the possible use of copper compounds, which are lethal to snails and other non-target animals. Anthelmintic treatment is a regular

practice in enzootic areas, but fails to eradicate the parasite. Allopathic anthelmintics are not completely effective against common flukes (5) and have serious disadvantages in some developing countries in addition to cost, risk of misuse leading to drug resistance, environmental pollution and food drug residues (6). Moreover, almost all adversely affect milk and meat production of animals during the course of their treatment, and even for long after their use (5).

Infestation of goat herds results in major health issues which lead to poor animal performance and economic loss. Currently, chemical anthelmintics are used to control gastrointestinal parasites. However, the increased resistance of parasites to anthelmintics will eventually result in a major crisis (7).

Concerning the relationship between bacteria and *Fasciola* spp., (8) observed the presence of large numbers of bacteria in the bile ducts of rats infected with *Fasciola hepatica* and their absence from uninfected animals suggests that the helminth in some way alters the conditions in the bile duct to favour bacterial growth in previously unfavourable environment. Indeed, significant changes have been found in the chemical characteristics of bile fluid from rats and cattle after liver fluke infection. After fluke infection the physico-chemical nature of the bile may have changed sufficiently to favour the multiplication of the particular bacteria. Kahan et al. 2008 (7) reported that, the bile duct is not normally inhabited by bacteria and the powerful detergent action of bile in disrupting certain types of bacteria is commonly used to differentiate the morphologically identical but harmless *Streptococcus viridans* from the virulent *S. pneumoniae* in the diagnosis of bacterial pneumonia. However bacterial invasion of both intestines and bile duct commonly accompanies biliary diseases and inflorescences of *Escherichia coli* and *Streptococcus faecalis* and other bacteria are not uncommon in the bile of patients with obstructive biliary disease Erlandsen and Chase 1972 and (10) reported that, there are two possible methods of introduction of the bacteria; one is direct migration when the fluke moves from the small intestine through the liver, while the other is migration of bacteria normally present in the small intestine into the bile duct after the establishment of the liver fluke.

Cheema and Myhammed 1980 (11) and (12) studied microorganisms associated with abscesses in sheep and goats. The bacteria isolated were *Escherichia coli*, *Staphylococcus epidermidis*, *Pasteurella* spp., *Streptococcus* spp., *Corynebacterium* spp., *Moraxella osloensis* and *Pseudomonas aeruginosa* from the sheep abscesses and *Staphylococcus epidermidis* from, *Peptostreptococcus anaerobius*, *Eubacterium tortuosum*, *Corynebacterium pyogenes*, *Pasteurella haemolytica*, *Pasteurella multocida* and *Corynebacterium pseudotuberculosis* from the goat liver abscess.

Foster 1984 (13) and (14) stated in the presence of fluke infection bacteria is thought to flourish due to changes in the biliary environment. This was confirmed in the early chronic phase, where a bacterial infection of the common bile duct was detected at 8 weeks post-infection in *F. hepatica* infected rats. They also stated, bile taken from rats infected with the liver fluke, *Fasciola hepatica* contained spiral bacteria, whereas bile from uninfected rats was free from spiral bacteria. The bacteria and its relationship to the bile duct epithelium and the liver fluke was studied with a combination of light microscopy, scanning and transmission electron microscopy. Its morphological characteristics suggest that the bacteria belong to the genus *Spirillum*. In contrast to many other co-infections of bacteria and helminths, the present one seems to be a fairly passive relationship so that neither the helminths nor the rat suffers from the presence of bacteria. Eguale and Abie 2003 (1), (15), (16) and (17) reported that, bacterial infection will often occur as a consequence of tissue damage, and the method of infection of many helminth parasites seems to be particularly likely to lead to the introduction of a secondary infection. Infested animals with *Fasciola* spp had suffered severe body weight loss and growth retardation. This economic loss can be due to anorexia caused in infected animals by restlessness. The bacteria will release toxins into the blood streamer resulting in what is known as black disease in sheep. Gonzalo-Orden et al. (18) reported introduction of bacteria into the biliary system during migration from the duodenum *Escherichia coli*, *Klebsiella pneumoniae* and *Enterococcus faecalis* from bile of infected sheep. Mas-Coma et al. 2000 (19) stated in the rats, the bacterobilia was higher when the number of parasites in the common bile duct were increased. Concerning fascioliasis, there is considerable tissue reaction and calcification in the bile ducts due to the presence of even a small number of flukes. Sayed et al. 2008 (20) observed that, invasion of the liver by migrating immature liver fluke damages the tissue and results in reduction of the oxygen tension (anaerobic condition), that allows the germination and proliferation of clostridial spores with release of its toxins and induce hepatocellular necrosis. William et al. 2008 (21) studied the fascioliasis and bacterobilia in experimentally and naturally infected animals, *E. coli* (50%), *Klebsiella pneumoniae* (30%), *Pseudomonas* spp (8%),

Proteus spp(5%) were isolated. Tehrani et al. 1998 (22) recorded that, the following bacteria were isolated: Corynebacterium spp. (52.4%), Pasteurella spp. (11.3%), Escherichia coli (11.1%), Pseudomonas aeruginosa (7.2%), staphylococcus spp. (5.3%). Borai et al. 2013 (23) stated that, bacteriological examination revealed that Staphylococcus aureus, Corynebacterium pyogenes and Escherichia coli were the most common aerobic bacteria while Clostridium perfringens and Fusobacterium necrophorum were the most common anaerobic bacteria isolated from the affected livers. Corynebacterium oviforme was isolated from six cases of sheep.

This study was designed to: -

Study the pathological hepatic lesions induced by bacteria in liver infested with Fasciola and to detect the relationship between these bacterial infections and liver fascioliasis. Also study, in vitro, the effects of some medicinal Herbal drug extracts (Calligonum comosum plant) as antimicrobial compound.

MATERIALS AND METHODS:

Bacterial isolation and characterization:

A loopful from each affected livers, bile duct and surface of Fasciola streaked onto the nutrient agar, MacConkey agar and blood agar plates, then incubated aerobically at 37 °C for 24 hrs. Bacterial isolates were identified morphologically, according (24).

Identification of bacteria species.

Identification of bacteria species were done by Baltimore Biology Laboratory (BBL) Crystal panel viewer of serial number 042611-007

Crystal Spec Nephelometer of serial number 11050002734

BBL Crystal mind software

BBL Crystal gram – positive & enteric / nonfermenter ID Kits, Cat number 245140 & 245000 respectively

BBL Oxidase Reagent Dropper Cat, number 261181 & BBL Indole Reagent. Dropper number 261185

Microbiological studies

Bile samples were simultaneous and examined according to (21), Fasciola worms were collected and counted under a dissecting microscope. First, the bile duct was examined for the presence of worms and stones, and then the rest of the organs were evaluated, the liver parenchyma, in particular (25).

Antimicrobial activity:

The antibacterial activity was evaluated by noting the zone of inhibition against the test organisms according to (26) and (27). Determination of minimal inhibitory concentration (MIC) of the extract was done according to (28) and (26).

Minimal bactericidal concentration (MBC)

The Minimum bactericidal concentration (MBC) of the plant extract was determined using the method described by (29).

Pathological and bacteriological studies on livers affected with fascioliasis in goats.

The lesion specimens were collected from four affected livers, and placed in sterile plastic containers and shipped to the laboratory, on ice, for bacterial culturing. In laboratory and under complete sterile condition in laminar flow cabinet samples were taken from three places (liver surface, bile duct and surface of adult Fasciola).

Identification of gram positive and negative bacterial isolates was done using standard procedures (24) and (30).

RESULTS AND DISCUSSION.

Types of bacteria isolated are shown in table (1, 2) and graph (1, 2, 3).

Table (1): Bacteria species isolated from different location of livers infected with *Fasciolagigantica*.

Location	Liver surface	Bile duct	Surface of <i>Fasciolagigantica</i>
Bacterial species isolated	<i>Stenotrophomonas Malyophilia</i>	<i>Aerococcus Urinae</i>	<i>Leifsonia aquatica</i>
	<i>Corynebacteriumpropinquum</i>	<i>Escherichia coli</i>	<i>Lactococcuslactis spp. cremoris</i>
	<i>Escherichia coli</i>		<i>Corynebacteriumrenale group</i>
			<i>Corynebacterium species</i>
			<i>Escherichia coli</i>

Table (2): Types and pathogeneses of the positive and negative gram microorganisms isolated from livers infested with fascioliasis.

Types of liver samples bacterial isolates	Pathogeneses
<u>Gram –positive</u> 1- <i>Leifsoniaaquatica</i>	Associated with the environment. Isolated occasionally in the clinical laboratory, but natural and pathogenic significances yet to be dined. Associated with endophthalmitis, meningitis in a child, and septicemia as well as urinary tract infection in a child.*
2- <i>Lactococcuslactis spp. cremoris</i>	With few case reports of being an opportunistic pathogen.*
3- <i>C. renale group</i>	Pathogenic veterinary bacterium that causes cystitis and pyelonephritis in cattle.*
4- <i>C. spp.</i>	Diphtheria some others animal.*
5- <i>C. propinquum</i>	Respiratory pathogen.*
6 - <i>Aerococcusurinae</i>	Predisposed to urinary tract infection.*
<u>Gram- negative</u> 1- <i>Stenotrophomonasmaltophilia</i>	Associated with human urinary tract and respiratory tract infections postoperative.*
2- <i>Escherichia coli</i>	Bacteremia and bacteria-related travelers diarrhea leading cause of neonatal meningitis and other infections including pneumonia.*

According to (31), (32).



Fig (1) Bacteria isolated from liver surface infected with *Fasciolagigantica* and cultivated on blood agar (A, B) and manconkey (c).

- A) *StenotrophomonasMalyophilia*
- B) *Corynebacteriumpropinquum*
- C) *Escherichia coli*

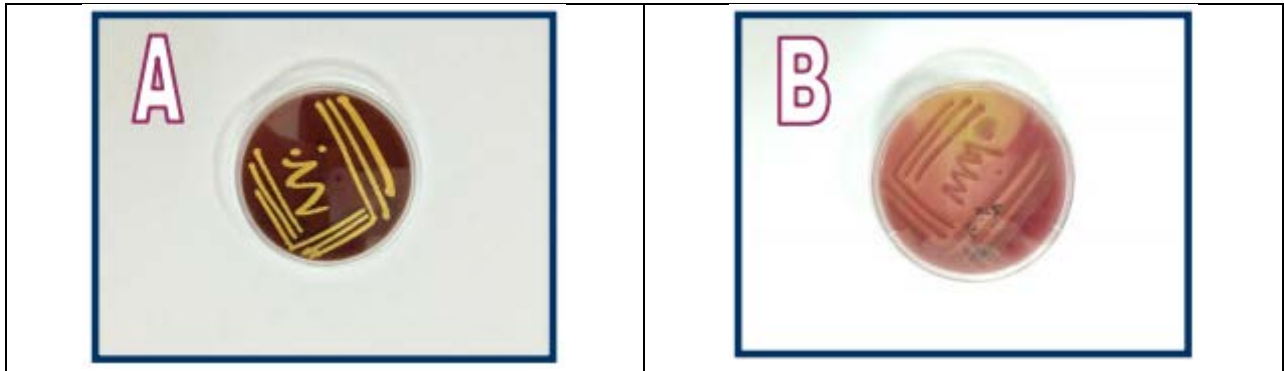


Fig (2) Bacteria isolated from bile duct infected with *Fasciolagigantica* and cultivated on blood agar(A) and manconkey(B).

- A) *AerococcusUrinae*
- B) *Escherichia coli*

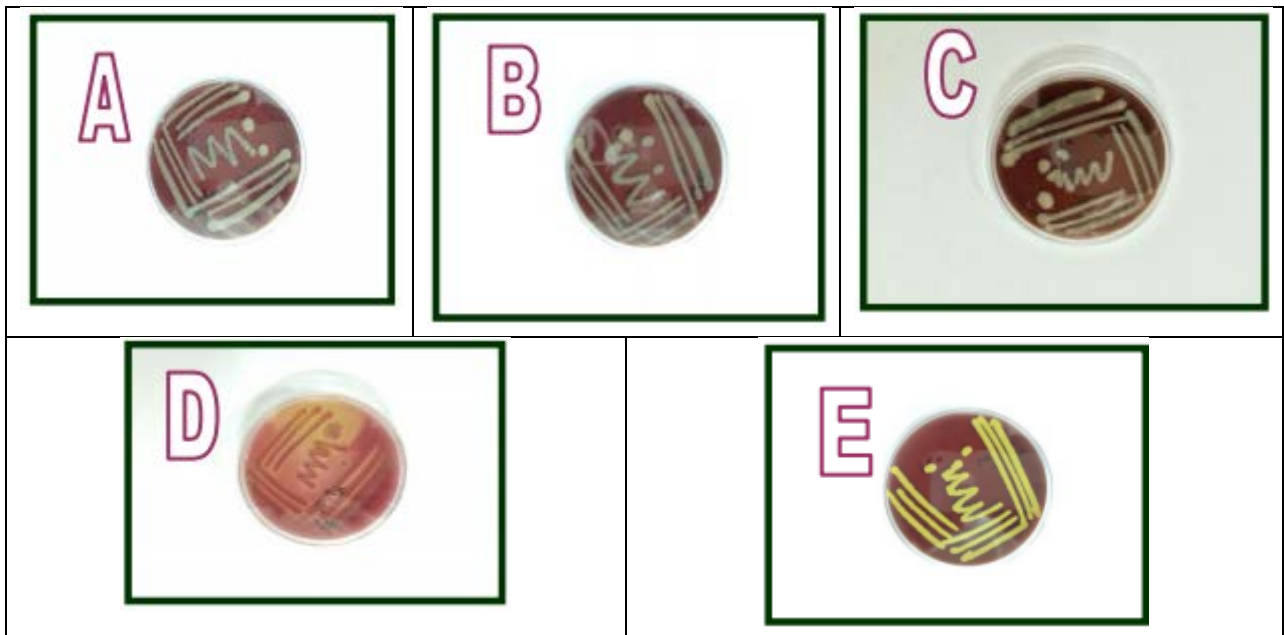


Fig (3) Bacteria isolated from surface of *Fasciolagigantica* and cultivated on blood agar (A, B, C, D) and maconkey (C)

- A) *Lactococcuslactis spp. (cremoris)*
- B) *Leifsoniaaquatica*
- C) *Corynebacteriumrenale*group
- D) *Corynebacterium*spp. E) *Escherichia coli*

DISCUSSION

Concerning pathological and bacteriological studies on livers affected with fascioliasis in goats.

Emerging evidence suggests a strong interaction between the gut microbiota and health and disease. The interactions of the gut microbiota and the liver have only recently been investigated in detail. The liver is receiving approximately 70% of its blood supply from the intestinal venous outflow, represents the first line of defense against gut-derived antigens and is equipped with a broad array of immune cells (macrophages, lymphocytes, natural killer cells, and dendritic cells) to accomplish this function. In the setting of tissue injury, whereby the liver is otherwise damaged (e.g., viral infection, toxin exposure, ischemic tissue damage, etc.), these same immune cell populations and their interactions with the infiltrating gut bacteria likely contribute to and promote these pathologies. Liver is

considered the most important organ for animal health production and reproduction. Many of the metabolic activities of the body occurred in the liver. Liver infection is an important disease that affects all kinds of meat producing animals, this lead to great losses to live-stock production and national income due to condemnation of great numbers of livers in the slaughter houses (33), (34) and (35).

Besides, infected liver constitutes a good media for bacterial multiplication, transportation of microorganisms with the parasites occurs during the different stages of its life cycle either outside or inside the animal body. Presence of both immature and mature flukes of *Fasciola* spp. cause acute and chronic hepatitis.

This study was carried out to detect the relationship between these bacterial infections and liver fascioliasis. The results of this study (Table 1) revealed that, the several types of bacteria which isolated from liver surface suffering from fascioliasis were *Stenotrophomonas malyophilia*, *Corynebacterium propinquum* and *Escherichia coli*. While *Aerococcus* spp. and *Escherichia coli* were isolated from the bile duct of the same infected liver. The results agreed with those obtained by (9), (1), (2), (13), (20) and (36), who reported that, there are 2 possible methods of introduction of the bacteria; one is direct migration when the fluke moves from the small intestine through the liver, while the other is migration of bacteria normally present in the small intestine into the bile duct after the establishment of the liver fluke. They stated also, bile bacteria and its relationship to the bile duct, due to changes in the biliary environment produced as a result of the fluke infection, these changes subsequently allow a multiplication of bacteria normally present in the uninfected animal.

Table (2) showed the predominant incidence of the positive and negative gram microorganisms isolated from livers infested with fascioliasis which were *Leifsonia aquatica*, *Lactococcus lactis* spp. *cremoris*, *Corynebacterium renale* group, *Corynebacterium* spp., *Corynebacterium propinquum*, *Aerococcus* spp., *Stenotrophomonas malyophilia* and *Escherichia coli*. Similar findings were described by (37), (38) and (39). Also, (40) who mentioned that, the most common bacteria isolated from the liver were *A. pyogenes* and anaerobic bacteria.

Bacteriological examination of samples with chronic fascioliasis showed the most frequently isolated aerobic or facultative organisms were *E. coli* and *Corynebacterium propinquum*. These results agreed with the previous findings of (41), (42) and (43) who found that *Fasciola* infestation plays an important role in the damages which were attributed to toxic environment created by the organisms in liver tissue. The presence of some members of family *Enterobacteriaceae* in combination with other *Clostridial* bacteria such as *C. perfringens* may be explained by (44) that infection with pathogenic *E. coli* may cause altering the mucous membrane of the intestine, enable the *clostridial* microorganisms to vegetate and release its toxins which absorbed through the damaged gut and reached to the blood circulation and then to the liver. On the other hand, (45) considered *E. coli* as an incidental pathogen which contaminated the animal tissues during preparation of the carcasses from faecal material, skin and hides. These results nearly coincide with that observed by (41), (46) and (47). Similarly (48) isolated *C. novyi* and *C. septicum* in incidences of 6% and 2% respectively.

From the achieved results, it is concluded that livers of goats showed a very high proportion of gross lesions rather than different pathological disorders. Also *Fasciola* worms may be incriminated in aiding bacterial infections specially

Stenotrophomonas malyophilia, *Corynebacterium propinquum*, *Aerococcus* spp., *Leifsonia aquatica*, *Lactococcus lactis* spp. *cremoris*, *Corynebacterium renale* group, *Corynebacterium* spp. and *E. coli* which lowering the hepatic viability. Generally, livers of slaughtered ruminant are considered as hazardous source of different mixed bacterial species. Moreover, their low value referred to high incidence of pathological lesions. During their migration phase through the abdominal cavity, liver parenchyma and bile ducts walls, young liver flukes may produce serious acute inflammatory tissue reaction, and blood loss owing to mechanical trauma and the young flukes then reach maturity in the bile ducts. These small liver flukes mainly produce chronic tissue reactions of the liver, such as fibrosis of small bile ducts, portal veins, and hepatic artery leading to biliary cirrhosis. Heavy infections may result in weight loss and emaciation and death of the host is mostly due to severe anemia and failure of liver function. Valero et al. 2006 (49) suggested an association between bacteribilia and both duration and intensity of parasitic infection. They added that the obstruction caused by advanced chronic fascioliasis in the animals may be related to biliary sepsis.

Grossly, post mortem examination of the livers infested with *Fasciola* in the present study, were hard, firm and tough in consistency with multiple, irregular pale brownish areas on the surface. Cut section of the liver revealed presence of large whitish areas of fibrosis. The affected ducts were enlarged, thickened, hard in consistency and in the form of cord like structures and protruded above the surface of the liver. In most cases, mature *Fasciola* worms were detected within the lumen of the affected bile ducts and bile stones. There are strong association between time of infection with *Fasciola*, number of flukes, and the risk of developing pigment stones in the main bile duct. Gallstone presence increased with infection time. The relative risk of gallstone disease increased when the number of flukes per animals increased. The *Fasciola* count in this study revealed more than 300 flukes, post mortem findings were attributed to the effect of toxic products elaborated by *Fasciola* worms and presence of mature worms within the lumen of intra hepatic bile ducts. These findings were coincided with (2), (16), (25), (19), (20), (49) and (50).

Therapeutic efficacy of ethanolic extract *C.comosum* extracts on different types of bacteria.

Bile is usually sterile and the important factors in maintaining its sterility are the choledochalsphincter, the bile flow, and its bacteriostatic properties. When there is obstruction and stagnation, bacteria gain access to the biliary system through either the papilla or the portal circulation.

The causes of biliary obstruction that predispose to bacterial cholangitis are myriad. Biliary parasites cause necrosis, inflammation, fibrosis, strictures, and bile ducts cholangiectasis by several mechanisms as a direct result of the irritating chemical composition of the parasite, parasitic secretions, or eggs physical obstruction of the bile ducts induction of biliary stones formation; and introduction of bacteria into the biliary system during migration from the duodenum. The main aim of this work is to investigate the relationship between presence of pathogenic bacteria in the bile versus fascioliasis using parasitological criteria: worm bacteriological bile culture and bacteriological fluke surface culture.

Concerning incidence of the positive and negative gram microorganisms isolated from livers infested with fascioliasis Table (1) showed that, the bacteriological bile culture revealed isolation of *Aerococcus Urinae* and *Escherichia coli*. Bacterial species isolated from liver surface were *Stenotrophomonas malyophilia*, *Corynebacterium propinquum* and *Escherichia coli*. While bacterial species isolated from *Fasciola gigantica* were *Leifsonia aquatica*, *Corynebacterium renale* group, *Corynebacterium* species, *Escherichia coli* and *Lactococcus lactis* spp. cremoris. All the isolated microorganisms are pathogenic and can be transmitted from animals to human causing very dangerous zoonotic diseases.

The present investigation evaluated the in-vitro antibacterial activity of the crude ethanolic extracts of *Calligonum comosum* 10% on some gram positive bacteria (*Corynebacterium propinquum*, *Aerococcus urinae*, *Leifsonia aquatica*, *Lactococcus lactis* spp.) cremoris, *Corynebacterium renale* group and *Corynebacterium* spp. and Gram negative bacteria (*Stenotrophomonas malyophilia* and *Escherichia coli*.)

Taking into consideration medicines from plant origin as an alternative form of health care is increasing because they are serving as promising sources of novel antibiotic prototypes (51). Plants are rich in a wide variety of secondary metabolites, such as tannins, terpenoids, alkaloids, and flavonoids, which have been found to have antimicrobial properties as anticancer agents, anti-diarrheal as well as antifungal activities (52) and (53).

In our current investigations we have found that the activity of ethanolic extract of *Calligonum comosum* 10% against the gram positive and Gram negative bacteria showed the highest zone of inhibition for gram negative and positive bacteria. These findings might be in well agreement with (54) who observed that the significant level of antibacterial activity was in a similar range of inhibition zone for the growth of the bacterium, *Listeria ivanovii* (9 – 18 mm) when laboratory tested by the agar well diffusion method. The authors also demonstrated that the most active plant part causing inhibition is the extract from the leaves. Thus; the study ascertains the value of plant parts used, which could be of considerable interest towards the development of new drugs. The observed differences in sensitivity the *Calligonum comosum* extract between gram positive and gram negative bacteria can probably be attributed to the structural and compositional variations in the nature of the cell wall between the two groups (55).

Results in current study were strongly agreed with results provided by (56), (57), (58) (59), (2), (3), (36) and (54).

Further in vivo studies are required to validate our findings and improve our knowledge on the potential of the *Calligonumcomosum* extract as antimicrobial in relation to its chemical composition.

Amazing and interesting to, isolated *Lactococcuslactis* spp.cremoris from the surface of *Fasciolagigantica*, collected from liver chronically suffered with *Fasciolagigantica*. Isolation of this microbe proved that the migration of bacteria can occur during the life cycle of *Fasciola*. However, it is a members of the genus *Lactococcus* and gram positive lactic acid bacteria.

These lactococci are found in various environments including plant surfaces (60) and (61), and recognized as pathogens in fish aquaculture (62). *Lactococcuslactis* is used worldwide in starter cultures for the manufacture of fermented dairy products, such as in the cheese-making process. *Lactococcuslactis* spp. can be found in various environments including animal sources, dairy products and silages (60), (61) and (63).

Lactococcuslactis has been observed at high numbers in the intestinal tract of the puffer fish *Takifuguniphobles* (marine fish). Additionally, the isolation of *L. lactis* strains from Amur Catfish *Silurusasotus* cultured in freshwater has been reported (64). Some strains of *L. lactis* have been isolated from intestinal tracts of several freshwater fish species. Mitchell 2002 (65) and (66) recorded that *L. lactis*spp.lactis may be isolated from the intestinal tracts of marine fish , and their excreta are continuously being introduced to the environment.

CONCLUSION

This study is of value in endemic areas, where possibility of bacteria co-infection with *Fasciola* is a common occurrence and highlights the importance of early chemotherapeutic intervention.

In the light of our present findings we can conclude that the ethanol leaf, root/bark extracts of *Calligonumcomosum* showed antibacterial effect in experimental models against *Corynebacteriumpropinquum*, *Aerococcusurinae*, *Leifsoniaaquatica*, *Lactococcuslactis* spp. cremoris, *Corynebacteriumrenale* group, *Corynebacterium* spp., and gram negative bacteria *Stenotrophomonasmaiophilia* and *Escherichia coli* which therefore offer a scientific basis for using this plant as a good source of traditional microbiological and anthelmintic references. Further studies are required to validate our findings and improve our knowledge on the potential of the *Calligonumcomosum* extract as antimicrobial in relation to its chemical composition. Isolation of *Lactococcuslactis* spp. cremoris may therefore have commercial application such as probiotics for aquaculture and the development of functional foods and novel food additives.

REFERENCES

- 1-Egual, T. and Abie, G. (2003): Some clinico-pathological observations of sheep artificially infected with *Fasciolagigantica*. *Bull. Anim. Health prod. Africa.*, 51(2):113-115.
- 2-Sohair, B. and Eman, N. (2009): Histopathological and bacteriological studies on livers affected with fascioliasis in cattle. *Egypt. J. Comp. Path. & Clinic. Path.*, 22:19-43
- 3-Nabila S. Degheidy and Jamila S. Al-Malki (2013): Incidence and evaluation of anthelmintic efficacy of *B.egyptiaca* and TCBZ on fasciolosis among goats in Taif, KSA. *Global Veterinaria*, 10(1):65-70.
- 4-Maqbool, A.; Hayat, C. S. and Tanver, A. (2004): Comparative efficacy of various indigenous and allopathic drugs against fasciolosis in buffaloes. *Vet. Arhiv.* 74: 107-114.
- 5-Brander, G. C.; Pugh, D. M. And Biwater, M. (1991): *Veterinary applied pharmacology and therapeutics*. 4th ed. Bailliere Tindall, London.
- 6-Hammond, J.A.; Fielding, D. and Bishop, S.C. (1997): Prospects for plant anthelmintics in tropical veterinary medicine *Vet. Res. Communic.*

- 7-Kahan, T.; Williams, S.; Mobley, R.; Ezenwa, I. and Peterson, E. (2008): The use of tanniferous plants to control infestations of *Haemonchus contortus* parasites in meat goats. University of Florida AN199
- 8-Haiba, M.H.; El-Rawii, K.A. and Osman, H.G. (1964): A comparative study on the levels of calcium inorganic P and MG, in the blood serum and bile of the normal healthy buffalo and those of buffaloes infested with liver fluke (*Fasciolagigantica*). *J. parasitenkunde.*, 23:527-531.
- 9-Erlandsen, S.L. and Chase, D.G. (1972): Paneth cell function: phagocytosis and intracellular digestion of intestinal microorganisms. *Journal of Ultrastructure Research*, 41:319-333.
- 10-Smith, H, A.; Jones, T.C. and Hunt, R.D. (1972): In veterinary pathology, 4th edn. Philadelphia; Lea and Febiger
- 11-Cheema, A.H. and Myhammed, S.I. (1980): Pathology of liver abscesses. *American Journal of Veterinary Research.*, 41-70.
- 12-Tadayon, R.A.; Cheema, A.H. and Muhammed, S.I. (1980): Microorganisms associated with abscesses of sheep and goats in soyth of Iran. *Am. J. Vet. Res.*, 41(5):798-802.
- 13-Foster, J.R. (1984): Bacterial infection of the common bile duct in chronic fasciolosis in the Rat. *J. Comp.Path.*, 94.
- 14-Kelly, W.R. (1985): The liver and biliary system in: pathology of domestic animals. Jubb K.V.F., Kennedy P.C. And Palmer N., 2:
- 15-Williams, A.M.; Fryer, J.L. and Collins, M.D. (1990): *Lactococcus piscium* sp. nov. *Lactococcus* sp from snid fish. *FEMS Microbiol Lett.*, 68:109-114.
- 16-Diehl, A. K.; Schwesinger, W. H.; Holleman, D.R.; Chapman, J.B. and Hurtin, W.E. (1995): Clinical correlates of gallstone composition: distinguishing pigment from cholesterol stone. *Am.J.Gastroenterol.*, 90:967-972
- 17-El-Azazy, O.M.E. (1995): Seasonal changes and inhibited development of the abomasal nematodes of sheep and goats in Saudi Arabia. *Vet.Parasitol.*, 58:91-98.
- 18-Gonzalo-Orden, M.; Millan, L. and Alvarez, M. (2003): Diagnostic imaging in sheep hepatic fasciolosis: Computer tomography and magnetic resonance findings. *Parasitol Res.*, 90: 64-359.
- 19-Mas-Coma, S.; Bargues, M.D.; Marty, A.M. and Neafie, R.C. (2000): Hepatic trematodiasis. *Hepatic trematodiasis. Helminthiasis, Armed forces institute of pathology and American Registry of pathology, Washington, DC*, 1:69-92.
- 20-Sayed, S. M.; Gehan, M.S. and Neveen, A.E. (2008): Clinicodiagnostic studies on hepatic affections of aged buffaloes. *Assiut Vet .Med.J.*, 54(117):310-328.
- 21-William, S.; Shalaby, M. and Ramzy, F. (2008): Fascioliasis and bacterobilia in experimentally and naturally infected Animals. *Parasitologists United Journal.*, 1(2):123-128.
- 22-Tehrani, A.; Javanbakht, J.; Hassan, M.; Zaman, M.; Rajabian, M.; Nagaraja, T.G. and Chengappa, M.M. (1998): Liver abscesses in feedlot cattle: a review. *J Anim Sci.*, 67:287-298
- 23-Borai, G.E.; Nagi, A.A.; Gab-Allah, M.S.; Abdel-Baset, I. and Moustafa, S.A. (2013): Comparative pathological studies on bacterial affections of liver in Farm Animals.
- 24-Quinn, P.; Markey, B.; Carter, M.; DONnelly, W.; Leonarrd, F. and Magnia, D. (2002): *Veterinary Microbiology and Microbial Diseases*. 1st ed. Published, Blackwell Science Ltd.

- 25-Valero, M.A.; Santana, M.; Morales, M.; Hernandez, J. and Mas-Coma, S. (2003): Risk of gallstone disease in advanced chronic phase of fasciolosis; an experimental study in a rat model. *J. Infect. Dis.*, 188:787-793.
- 26-Eloff, J.N. (1998): A sensitive and quick microplate method to determine the minimal inhibitory concentration of plant extracts for bacteria. *Planta Medica.*, 64 (8): 711 – 713.
- 27-Andrews, J.M. (2001): Determination of minimum inhibitory concentrations. *Journal of Antimicrobial Chemotherapy.*, 48 (1): 5 – 16.
- 28-Oboh, I.E.; Akerele, J.O. and Obasuyi, O. (2007): Antimicrobial Activity of the Ethanol Extract of the Aerial Parts of *Sida acutangula* (malvaceae). *Tropical Journal of Pharmaceutical Research*, 6 (4): 809-813.
- 29-Igbinosa, O. O.; Igbinosa, E.O. and Aiyegoro, A.O. (2009): Antimicrobial activity and phytochemical screening of stem bark extracts from *Jatropha curcas* (Linn). *African Journal of Pharmacy and Pharmacology*. 3 (2): 058 – 062.
- 30-Konoman, E.W.; Paul, C.; Al-len, W. and Washington, C. (1992): Color atlas and textbook of diagnostic microbiology. 4th. J.B. Lippincott Company Philadelphia.
- 31-Baron, E.J. and Tenenbaum, S.M. (1990): *Bailey and Scott's diagnostic microbiology*, 8th Ed. C.V. Mosby Co., St. Louis, Mo.
- 32-Holt, J.G.; Krieg, N.R.; Sneath, P.H.A.; Staley, J.T.; Williams, S.T. (1994): *Bergey's manual of determinative bacteriology*, 9th Ed.
- 33-Foster, L. and Woods, W. (1970): Influence of liver abscesses on animal performance. *J. Anim. Sci.*, 31:44.
- 34-Tomate, H. (1973): High Incidence of ruminal lesions and liver abscess in prefecture. " *Tohoku, J. Agr. Res.*, 23, 184-199.
- 35-Nabila S. Degheidy and Jamila S. Al-Malki (2012): Epidemiological Studies of Fasciolosis in Human and Animals at Taif, Saudi Arabia. *World Applied Sciences Journal*, 19 (8): 1099-1104.
- 36-Dalal, H.M. Alkhalifah (2013): In-vitro antibacterial activity of ethanol extract of *Calligonum comosum* plant against four human pathogens in Saudi Arabia. *International Journal of Plant, Animal and Environmental Sciences*, 3(4):170-175.
- 37-Hanelein, G.F.W. and Abdellatif, M.A. (2003): Trends in small ruminant husbandry and nutrition and specific reference to Egypt. *Small ruminant res.*, 15:185-200.
- 38-Rosa, J.S.; Johnson, E.H.; Alves, F.S. and Santos, F.L. (1989): A Retrospective study of hepatic abscesses in goats: pathological and microbiological findings. *Br. Vet. J.*, 37.
- 39-El-Sayed, M. H. and Allam, A. F. (1997): Effect of triclabendazole on the tegument of *Schistosoma mansoni* a scanning electron microscope study. *Journal of Egyptian Society of parasitology*. 27, 143-152
- 40-Dore, E.; Fecteau, G.; Helie, P. and Francoz, D. (2007): Liver abscesses in Holstein dairy cattle." *Journal of Veterinary Internal Medicine*, 21 (4):853-856.
- 41-Zaki, O.A. (1956): The incidence of Salmonella infection in Camels. *J. Egypt. Publ. Health.*, 2:75-79
- 42-Samad, M.A. and Haque, M.E. (1987): Clinical occurrence of infectious necrotic hepatitis in cattle of Bangladesh. *Indian J. Anim. Health.*, 26(1):63-64.
- 43-Srokina, I. B. (1987): On the pathogenesis of necrotic hepatitis in fascioliasis." *Vet. (Moscow)*, 7:55-56

- 44-Wernery, U. (1992): The prevalence of Salmonella infections in Camels (*Camelus dromedarius*) in the United Arab Emirates. *Br.Vet.J.* 148(5):445-450.
- 45-Morshady, A.; Ammar, A.; Saleh, R. and Naenaesy, E. (1991): Microbial flora of camel liver." *Zagazig Vet. J.*, 19(4):940-944.
- 46-Darwish, M.M.F. (1996): Pathological studies on some liver affections in camels. Ph. D. Thesis, (Pathology Dept.) Fac. Vet. Med., Cairo University
- 47-Mohamed, H. M.; Thoria, I. EL Saied and Udall, H. Attia (1997): Pathologic studies on some liver affections in camels at Sharkia Governorate." *Egypt. J. Comp. Path. And Clin. Pathol.*, Vol 10, No.2., 17-27.
- 48-Leloglu, S. (1972): A comparative study of livers with or without liver-flukes (*Fasciola hepatica*) for clostridial infections by means of culture and by fluorescent antibody technique." *Turk-Veteriner-Hekimleri- Dergisi.*, 42(718): 19-22.
- 49-Valero, M.A; Navarro, M.; Garcia-bodelon, M.A.; Marcilla, A.; Morales, M.; Mengual, P. and Mas-Coma, S. (2006): High risk of bacterobilia in advanced experimental chronic fasciolosis. *Acta Tropica.*, 100:17-23.
- 50-Mahmoud, A. Z.; Uoussef M. S. and Ibrahim, M. K. (1989): "Pathological studies on some liver affections in camels. 1. Parasitic hepatitis (Disomisaia)." *Egypt. J. Comp. Pathol. and Clin. Pathol.*; 2(1):94- 106.
- 51-Koduru S, Grierson DS, Afolayan AJ 2006. Antimicrobial activity of *Solanum aculeastrum*. *Pharmaceutical Biotechnology* 44: 283 – 86.
- 52-Lucy, H. and Dasilva, E.J. (1999): Medicinal plants: Are-emerging health and electronic. *Journal of Biotechnology.*, 56-70.
- 53-Rauha JP, Remes S, Heinonen M, Hopia A, Kähkönen M, Kujala T, Pihlaja K, Vuorela H, Vuorela P (2000). Antimicrobial effects of Finnish plant extracts containing flavonoids and other phenolic compounds. *International Journal of Food Microbiology* 6 (1): 3 – 12.
- 54-Riadh H, Imen F, Abdelmajid Z, Sinda F. (2011): Detection and extraction of Anti-Listerial compounds from *Calligonum comosum*, a medicinal plant from arid region of Tunisia. *African Journal of Traditional Complementary and Alternative Medicine* 8 (3): 322 – 327.
- 55-Lambert P.A. (2002): Cellular impermeability and uptake of biocides and antibiotics in Gram-positive bacteria and mycobacteria. *Journal of Applied Microbiology* 92: 46 – 54. 56-Mahady, G.B. (2005): Medicinal plants for the prevention and treatment of bacterial infections. *Curr Pharm Des.*, 11(19): 2405-27
- 57-Badria, F.A.; Ameen, M. and Akl, M.R. (2007): Evaluation of cytotoxic compounds from *Calligonum comosum* L. growing in Egypt. *Z Naturforsch C*, 62(9-10): 656-60.
- 58-Bakray, F.A. (2009): Use of Some Plant Extracts to Control *Biomphalaria alexandrina* Snails with Emphasis on Some Biological Effects. *World Applied Sciences Journal*, 6(10): 1335-1345.
- 59-Tariq, K.A.; Chisti, M.Z.; Ahmed, F. and Shawl, S. (2009): Anthelmintic activity of extracts of *Artemisia afra* against ovine nematodes. *Vet. Parasitol.*, 160(1-2):8-38.
- 60-Ulrich, A. and Muller, T. (1999): Heterogeneity of plant-associated streptococci as characterized by phenotypic features and restriction analysis of PCR-amplified 16S rRNA. *J Appl. Microbiol.*, 84:293-303.
- 61-Salama, M.S.; Musafija-Jeknic, T.; Sandine, W.E. and Giovannoni, S.J. (1995) :An ecological study of lactic acid bacteria: isolation of new strains of *Lactococcus* including *Lactococcus lactis* subsp. *cremoris*. *J. Dairy Sci* 78, 1004–1017.

62-Cadmus, S.I.B. and Adesokan, H.K. (2009): Causes and implication of bovine organs/ offal condemnation in some abattoirs in western Nigeria. *Top. Amin. Health Porod.*, 4:1455-1463

63-Ennahar, S., Cai, Y. and Fujita, Y. (2003): Phylogenetic diversity of lactic acid bacteria associated with paddy rice silage as determined by 16S ribosomal DNA analysis. *Appl Environ Microbiol* 69, 444–451.

64-Sugita, H.; Ohta, K.; Kuruma, A. and Sagesakam T. (2007): An antibacterial effect of *Lactococcus lactis* isolated from the intestinal tract of the Amur catfish, *Silurus asotus* Linnaeus. *Aquac Res.*, 38:1002-1004.

65-Mitchell, G. (2002): Update on fasciolosis in cattle and sheep. *In practice.*, 24:378-385.

66-Itoi, S., Abe, T., Washio, S., Ikuno, E., Kanomata, Y. and Sugita, H. (2008) Isolation of halotolerant *Lactococcus lactis* subsp. *lactis* from intestinal tract of coastal fish. *Int J Food Microbiol* 121, 116–121.